

# THE TOXIC EXTRACTIVES FROM *WEDELIA ASPERRIMA*, III.<sup>1</sup> STRUCTURES OF TWO NATURALLY OCCURRING RHAMNOSYL ANALOGUES OF WEDELOSIDE

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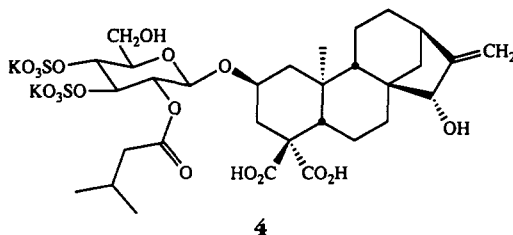
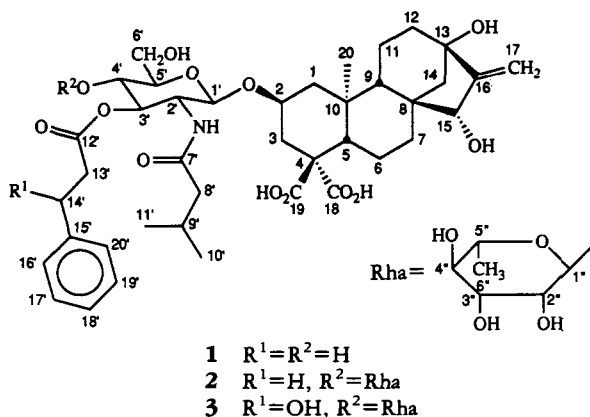
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**ABSTRACT.**—The structures of two new toxic 4'-*O*-rhamnosyl analogues of wedeloside [1], from *Wedelia asperima*, have been determined by a combination of chemical, nmr, and mass spectrometric analysis.

In previous papers, we described the structural elucidation of the diterpene aminoglycoside wedeloside [1], the major toxic constituent of *Wedelia asperima* Benth. (Compositae) (1,2). Subsequent biochemical studies (3) have shown wedeloside to be a powerful inhibitor of ADP/ATP transport across the mitochondrial membrane, with a binding affinity to the carrier protein comparable to that of the related diterpene glycoside, carboxyatractyloside [4] (4). We now report the structures of two additional toxic components 2 and 3 from *W. asperima*.

## RESULTS AND DISCUSSION

For the structure of *O*- $\alpha$ -L-rhamnosyl-(1'' $\rightarrow$ 4')-wedeloside [2], the fab mass spectrum of 2 showed an [MH]<sup>+</sup> ion at *m/z* 904 and an [MNa]<sup>+</sup> ion at *m/z* 926, which mass-



<sup>1</sup>For Part II, see Lewis *et al.* (2).

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measured for  $C_{46}H_{66}NO_{17}$  and  $C_{46}H_{65}NO_{17}Na$ , respectively, corresponding to wedeloside [**1**] plus an additional  $C_6H_{10}O_4$  unit. After permethylation of **2**, the eims of the product **5** showed an  $[M]^+$  at  $m/z$  911 ( $C_{47}H_{77}NO_{16}$ ) with prominent ions at  $m/z$  476 ( $C_{23}H_{42}NO_9$ ), 419 ( $C_{24}H_{35}O_6$ ), and 189 ( $C_9H_{17}O_4$ ) corresponding to the glycoside ( $[G]^+$ ), aglycone ( $[A]^+$ ), and terminal sugar ( $[S]^+$ ) fragments (Table 1). As was the case with wedeloside [**1**] (1), the 3-phenylpropanoyl group was lost from **2** during the permethylation procedure. On pertrideuteromethylation, the  $[M]^+$ ,  $[G]^+$ ,  $[A]^+$ , and  $[S]^+$  ions shifted to  $m/z$  941 ( $10 \times Me$ ), 494 ( $6 \times Me$ ), 431 ( $4 \times Me$ ), and 198 ( $3 \times Me$ ), respectively, in accordance with the expected number of active hydrogens (OH, NH) in the different portions of the molecule.

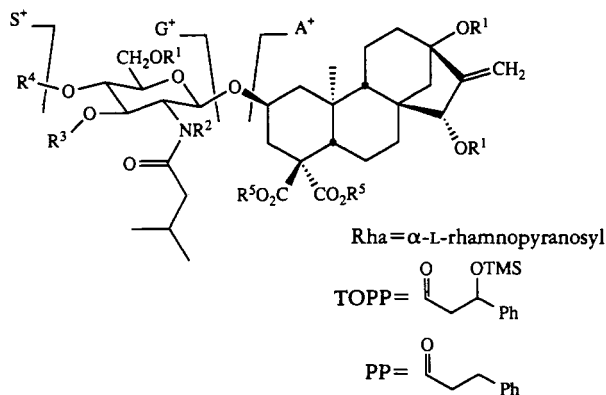
TABLE 1. Structurally Significant Ions ( $m/z$ ) in the Mass Spectra of Derivatized Toxins 1-3.

Compound	Derivative	$[M]^+$	$[A]^+$	$[G]^+$	$[S]^+$
<b>5</b> . . . . .	per Me <b>2</b>	911	419	476	189
<b>6</b> . . . . .	per TMSi <b>3</b>	1567	651	900	363
<b>7</b> . . . . .	per TMSi <b>1</b> <sup>a</sup>	1189	651	522	—

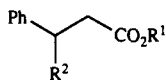
<sup>a</sup>Data are from Lewis *et al.* (2).

Acid hydrolysis of **2** in aqueous MeOH followed by trimethylsilylation and subsequent gc-ms of the product mixture showed the presence of TMSi-3-phenylpropanoate [**9**] (1), the tri-TMSi derivative **11** of methyl 2-deoxy-2-(3-methyl-1-oxobutyl)amino- $\alpha$ -D-glucopyranoside (1), and a 6-deoxyhexose derivative. The latter was compared with standard TMSi-6-deoxyhexoses and showed an identical mass spectrum and gc retention time to those of tetra-TMSi-L-rhamnopyranoside [**13**].

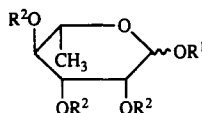
Methanolysis of **5**, the permethyl derivative of **2**, gave three major products. One was a compound with  $[M]^+$  at  $m/z$  436 and diagnostic fragment ions at  $m/z$  128, 145, and 165, confirming its structural identity with 13,15,18,19-tetra-O-methylwedeligenin [**16**] (2). The second was a compound with  $[M]^+$  at  $m/z$  319 corresponding to a tetramethyl derivative of the de-esterified 2-deoxy-2-acylaminoglucoside unit in **2** (i.e., structure **12**). The location of the free hydroxy substituent at C-4 in this compound (corresponding to the position of attachment of the rhamnopyranosyl moiety in **2**)



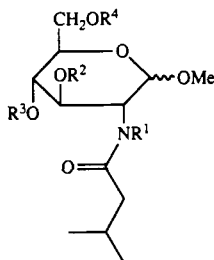
- 5**  $R^1=R^2=R^3=R^5=Me$ ,  $R^4=Rha(Me)_3$   
**6**  $R^1=R^5=TMSi$ ,  $R^2=H$ ,  $R^3=TOPP$ ,  $R^4=Rha(TMSi)_3$   
**7**  $R^1=R^4=R^5=TMSi$ ,  $R^2=H$ ,  $R^3=PP$   
**8**  $R^1=R^2=R^3=H$ ,  $R^4=Rha$ ,  $R^5=Me$



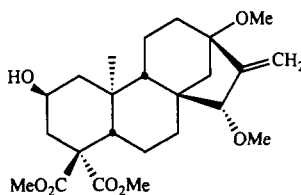
- 9**  $R^1 = \text{TMSi}, R^2 = \text{H}$   
**10**  $R^1 = \text{Me}, R^2 = \text{OH}$



- 13**  $R^1 = R^2 = \text{TMSi}$   
**14**  $R^1 = R^2 = \text{Me}$   
**15**  $R^1 = \text{Me}, R^2 = \text{H}$



- 11**  $R^1 = \text{H}, R^2 = R^3 = R^4 = \text{TMSi}$   
**12**  $R^1 = R^2 = R^4 = \text{Me}, R^3 = \text{H}$



**16**

could be deduced from the presence of well documented cleavage ions at  $m/z$  87, 170, and 171 in its mass spectrum (5). The third was a compound corresponding to a tetra-*O*-methyl-6-deoxyhexose, with structure **14**.

The basic skeletal structure of **2** was initially deduced by comparison of its  $^{13}\text{C}$ -nmr spectrum with that of wedeloside [**1**] (6) (Table 2). With the exception of C-4', the position of attachment of the rhamnosyl moiety, which showed the expected downfield shift of ca. 7 ppm, and the 3'- and 5'-carbons which exhibited small (2–3 ppm) upfield shifts, the chemical shift values for the carbon atoms of the aglycone (C-1 to C-20) and acylaminoglucoside (C-1' to C-20') moieties were directly comparable. The remaining six carbon resonances C-1'' to C-6'' corresponded to the values reported for a 1- $\alpha$ -linked L-rhamnopyranosyl residue (7). Verification of the structure of **2** was obtained using various 2D nmr experiments (COSY, long-range COSY, HETCOR, long-range HETCOR, and phase-sensitive double-quantum-filtered COSY). These experiments allowed direct correlations between geminally and vicinally coupled and long-range coupled protons as well as analogous proton-to-carbon correlations yielding complete and unambiguous chemical shift assignments. Most notable were the  $^{13}\text{C}$  chemical shift assignments of the glycosidic carbons and C-2. Severe overlap in the  $^1\text{H}$ -nmr spectrum of the protons associated with these carbons and the proximity of their  $^{13}\text{C}$  chemical shifts necessitated the use of these nmr spectral techniques. The quaternary carbons, as well as the C-5 and C-9 methines, were distinguished via their long-range  $^1\text{H}$ - $^{13}\text{C}$  couplings to their neighboring methylenes.

In the  $^1\text{H}$ -nmr spectrum of **5**, the two C-17 olefinic methylene protons were present at  $\delta$  5.13 and 5.24, while the anomeric protons on C-1' and C-1'' appeared at  $\delta$  4.6 ( $J = 8$  Hz) and  $\delta$  4.99 ( $J = 1$  Hz), respectively, the former supporting the assignment of a 1- $\beta$ -linkage for the 2-deoxy-2-acylaminoglucosyl group (1).

The fab spectrum of **3** showed a weak  $[\text{MH}]^+$  ion at  $m/z$  920 together with a stronger  $[\text{MNa}]^+$  ion at  $m/z$  942 ( $\text{C}_{46}\text{H}_{65}\text{NO}_{19}\text{Na}$ ) and ions for  $\text{CO}_2$  loss at  $m/z$  876 and 898, respectively. The per-TMSi derivative of **3** exhibited, in its eims, an  $[\text{M}]^+$  at  $m/z$  1567 and an  $[\text{A}]^+$  ion at  $m/z$  651, indicating the presence of the same aglycone moiety as in **7**, the per-TMSi derivative of wedeloside [**1**] (Table 1). Further, the  $[\text{S}]^+$  ion at  $m/z$  363 suggested that compound **3** also possessed a rhamnose sugar but differed from

TABLE 2.  $^{13}\text{C}$  Chemical Shift Assignments<sup>a</sup> for Compounds **1**, **2**, and **3**.

Carbon	<b>1</b> <sup>b</sup>	<b>2</b>	<b>3</b>
C-1	48.1	47.2	46.4
C-2	73.5	75.3	73.3
C-3	41.0	42.9	41.2
C-4	59.0	61.0	59.1
C-5	49.8	52.2	51.7
C-6	24.0	23.4	24.0
C-7	35.8	35.1	35.9
C-8	46.7	47.2	46.9
C-9	53.5	51.9	53.8
C-10	40.0	40.4	41.2
C-11	20.9	20.2	21.0
C-12	41.0	40.5	40.2
C-13	79.5	79.8	79.7
C-14	43.9	45.8	45.4
C-15	82.2	81.5	82.4
C-16	160.4	159.0	160.9
C-17	108.9	109.3	108.8
C-18	175.5	179.0	175.8
C-19	175.5	179.4	175.3
C-20	17.6	17.6	17.7
C-1'	101.2	99.7	101.0
C-2'	55.2	54.7	55.7
C-3'	77.0	75.4	76.7
C-4'	69.6	76.5	76.3
C-5'	77.4	74.7	75.9
C-6'	62.2	61.0	62.0
C-7'	175.5	177.3	175.3
C-8'	46.7	46.2	46.7
C-9'	27.2	26.7	27.2
C-10'	22.9	22.3	23.0
C-11'	22.9	22.4	23.0
C-12'	174.2	175.4	172.1
C-13'	31.5	35.8	48.2
C-14'	36.6	38.9	71.2
C-15'	141.7	141.3	145.2
C-16', -20'	129.2	129.1	127.1
C-17', -19'	129.2	129.7	129.4
C-18'	127.1	124.1	128.6
C-1''	—	102.0	102.8
C-2''	—	70.7	72.0
C-3''	—	71.2	72.4
C-4''	—	72.7	73.9
C-5''	—	70.3	70.8
C-6''	—	17.6	18.1

<sup>a</sup> $\delta$ , ppm from TMS, in  $\text{CD}_3\text{OD}$  (**1** and **3**) and  $\text{D}_2\text{O}$  (**2**).<sup>b</sup>Data are from Lewis and MacLeod (6).

**2** in having an extra hydroxy substituent located on the aminoglucoside moiety. The presence in the spectrum of an ion at  $m/z$  179 ( $\text{C}_7\text{H}_6\text{OTMSi}$ ) placed the hydroxy group on either the phenyl ring or C-3 of the phenylpropanoate group. Treatment of **3** with  $\text{CH}_2\text{N}_2$  followed by  $\text{NaOMe/MeOH}$  yielded, after hplc purification, methyl 3-hydroxy-3-phenylpropanoate [**10**], identical by ms and  $^1\text{H}$  nmr to an authentic racemic sample, while its  $[\alpha]_D$  value established the absolute stereochemistry of the chiral center as *S* (**8**). The residual compound **8** on methanolysis gave 1-*O*-methyl  $\alpha$ -L-rhamnopy-

ranoside [15], which had a  $^{13}\text{C}$ -nmr spectrum and optical rotation identical to authentic 1-*O*-methyl  $\alpha$ -L-rhamnopyranoside (7,9).

The  $^{13}\text{C}$ -nmr spectrum of **3** (Table 2) provided confirmation of its assigned structure. Apart from the carbon atoms at the 13', 14', and 15' positions, which showed the expected downfield shifts compared to those of **2** due to the additional hydroxy substituent on C-14', the spectra of **2** and **3** were comparable, any slight differences being attributable to solvent effects (MeOH vs.  $\text{H}_2\text{O}$ ).

## EXPERIMENTAL

**GENERAL EXPERIMENTAL PROCEDURES.**—Partial details of these have been reported previously (1). Additionally, some gc-ms were recorded on a VG7070F mass spectrometer equipped with a Varian 1400 gc using a 2 m  $\times$  2 mm glass column packed with 2% OV-17; some  $^1\text{H}$ - and  $^{13}\text{C}$ -nmr spectra were run on either a JEOL FX-200 or a Bruker HFX270 spectrometer. The COSY, long range COSY, HETCOR and long-range HETCOR experiments were carried out on a Varian VXR-300S spectrometer, while the phase sensitive double-quantum-filtered COSY and the long range version of the same experiment were performed on a Varian VXR-500 nmr spectrometer. Optical rotations were measured using a Perkin-Elmer 241 spectropolarimeter. Fabms accurate mass measurements were obtained on a VG ZAB-SEQ mass spectrometer using glycerol as matrix and polyethylene glycol as internal reference.

**ISOLATION OF 2 AND 3.**—The extraction and isolation procedure for **2** and **3** from *W. asperima* follows that reported for wedeloside [1] (1) up to the point of elution of the toxic fractions from a column of High Flow Super Cell. The dried fraction containing **2** and **3** was dissolved in MeOH- $\text{H}_2\text{O}$ -HOAc (40:60:1) saturated with 1,2-dichloroethane and applied to a column of micronized XAD-2 resin (50 g). Elution with the same solvent mixture (500 ml) was followed by stepwise elution with solvent mixtures (100 ml each) containing an increasing proportion of MeOH to  $\text{H}_2\text{O}$  but maintaining the same proportion of HOAc. Fractions from the column were monitored by tlc and after drying provided **2** and **3** as amorphous powders (60 mg and 200 mg, respectively). Compound **2** was further purified by chromatography on a Sephadex LH-20 column (2 m  $\times$  1 cm) using MeOH as eluent followed by a second chromatographic step on a Sephadex G-10 column (1 m  $\times$  1 cm) again using MeOH as eluent.

**Compound 2.**— $[\alpha]_{\text{D}}^{20} -34^\circ$  ( $c = 1.5$ , MeOH); fabms  $m/z$   $[\text{MH}]^+$  904.4410 (calcd for  $\text{C}_{46}\text{H}_{66}\text{NO}_{17}$ , 904.4331),  $[\text{MNa}]^+$  926.4140 (calcd for  $\text{C}_{46}\text{H}_{65}\text{NO}_{17}\text{Na}$ , 926.4150);  $^1\text{H}$  nmr  $\delta$  ( $\text{D}_2\text{O}$ , 100 MHz) 1.36 (d,  $J = 6$  Hz, Me), 5.32 and 5.41 (both bs,  $=\text{CH}_2$ ), 7.63 (bs,  $\text{C}_6\text{H}_5$ );  $^{13}\text{C}$  nmr ( $\text{D}_2\text{O}$ ) see Table 2.

**Compound 3.**—Fabms  $[\text{MNa}]^+$  942.416 (calcd for  $\text{C}_{46}\text{H}_{65}\text{NO}_{18}\text{Na}$ , 942.410);  $^{13}\text{C}$  nmr see Table 2.

**TOXICITY TESTING.**—This was carried out as previously reported (10). Both compounds **2** and **3** were toxic to mice and rats down to the 1 mg/kg level.

**PERMETHYLATION AND PERTRIDEUTEROMETHYLATION OF 2.**—Compound **2** (9.8 mg) was permethylated according to the procedure described previously for wedeloside (1). The permethylated derivative was purified by preparative tlc on silica  $[\text{MeCN}-\text{C}_6\text{H}_6, (40:60), R_f 0.39]$ , giving **5** (6 mg). Using the same procedure, but substituting  $\text{CD}_3\text{I}$ , the pertrideuteromethylated derivative of **2** was also prepared.

**Compound 5.**— $^1\text{H}$  nmr  $\delta$  ( $\text{CDCl}_3$ , 100 MHz) 4.60 (d,  $J = 8$  Hz, anomeric H), 4.99 (bs,  $J \sim 1$  Hz, anomeric H), 5.13 and 5.24 (both bs,  $=\text{CH}_2$ );  $^{13}\text{C}$  nmr ( $\text{CDCl}_3$ , 15 MHz) 16.7, 17.8, 20.4, 22.9, 25.2, 28.3, 34.8, 36.8, 39.5, 39.9, 42.2, 43.7, 46.5, 47.2, 50.3, 50.7, 51.1, 52.1, 53.0, 57.2, 58.0, 58.9, 59.5, 60.8, 62.8, 68.5, 71.5, 72.2, 77.9, 80.6, 81.1, 82.3, 84.6, 90.4, 98.0, 98.6, 109.3, 151.8, 172.5, 174.4; eims  $m/z$   $[\text{M}]^+$  911.5242 (calcd for  $\text{C}_{47}\text{H}_{77}\text{NO}_{16}$ , 911.5276), 896, 880, 706, 560, 476 (found 476.2850, calcd for  $\text{C}_{23}\text{H}_{42}\text{NO}_9$ , 476.2860), 444, 419 (found 419.2433, calcd for  $\text{C}_{24}\text{H}_{35}\text{O}_6$ , 419.2434), 387, 224, 189 (found 189.1128, calcd for  $\text{C}_9\text{H}_{17}\text{O}_4$ , 189.1127), 171, 165, 128.

**Pertrideuteromethylated 2.**—Eims  $m/z$   $[\text{M}]^+$  941, 926, 907, 727, 575, 494, 459, 431, 396, 230, 198, 177, 171, 134.

**HYDROLYSIS OF 2.**—Compound **2** (1.0 mg) in a mixture of MeOH (1 ml),  $\text{H}_2\text{O}$  (0.5 ml), and 4 M HCl (0.5 ml) was heated in a sealed tube at  $90^\circ$  for 2 h. After evaporation to dryness, the residue was silylated with TRISIL and analyzed by gc-ms (100–250 $^\circ$ ,  $\Delta 10$ ). Three major hydrolysis products were identified: TMSi-3-phenylpropanoate [9], by comparison of its ms and retention time with the TMSi derivative of an authentic sample (Aldrich); compound **11**, by comparison with its published mass spectrum (1); and tetra-TMSi-rhamnose, by comparison of its ms and retention time with the TMSi derivative of an authentic sample (Sigma).

**METHANOLYSIS OF 5.**—The permethylated derivative **5** (1.0 mg) was dissolved in MeOH (1 ml) and heated at  $60^\circ$  over Dowex 50W resin ( $\text{H}^+$  form) for 2 h. An aliquot, analyzed directly by gc-ms (100–

200°, Δ10), showed three major peaks: compound **16** ( $[M]^+$   $m/z$  436) identified as 13,15,18,19-tetra-*O*-methylwedeligenin [**16**] by comparison with its published mass spectrum (2); compound **12**, *eims m/z* (rel. int.)  $[M]^+$  319 (0.5), 304 (1.5), 288 (1), 287 (2), 274 (4), 272 (4), 244 (10), 224 (20), 171 (21), 170 (27), 141 (30), 140 (79), 117 (100), 87 (>100); and compound **14**, identified by comparison of its *ms* and retention time with those of authentic tetra-*O*-methyl rhamnose.

**SILYLATION OF 3.**—Compound **3** (1.0 mg) was suspended in dry MeCN (10 μl) and treated with BSTFA-TMCS (9:1) (50 μl) at 70° for 20 min to give the per-TMSi derivative **6**. An aliquot was subjected to direct probe *eims*. For significant ions, see Table 1. Accurate mass measurement on  $m/z$  179 gave 179.0894, C<sub>10</sub>H<sub>15</sub>O<sub>5</sub>Si requires 179.0892.

**HYDROLYSIS OF 3.**—Compound **3** (40 mg) in MeOH (2 ml) was first treated with CH<sub>2</sub>N<sub>2</sub> in Et<sub>2</sub>O, followed by addition of a small amount of sodium to the MeOH solution after removal of the Et<sub>2</sub>O. After 18 h at room temperature, the solvent was removed and the residue triturated with CHCl<sub>3</sub>. Hplc of the CHCl<sub>3</sub> washings on a Zorbax SIL column (25 cm × 4.6 mm) using CH<sub>2</sub>Cl<sub>2</sub>-MeCN (9:1) as eluent afforded (*S*)-methyl 3-hydroxy-3-phenylpropanoate [**10**] (2.6 mg),  $[\alpha]^{25}_D -15^\circ$  ( $c = 1.3$ , EtOH) [lit. (8) (*R*)-form +18.3°]. <sup>1</sup>H-, <sup>13</sup>C-nmr, and mass spectra were identical to an authentic sample of racemic **10**.

The residue from alkaline hydrolysis was taken up in MeOH (2 ml). Dowex 50W resin (H<sup>+</sup> form) was added and the mixture was heated at 60° for 2 h. After filtration and removal of solvent, the hydrolysate was chromatographed [plc; EtOAc-MeCN (9:1)] giving (*R*, 0.23) 1-*O*-methyl α-L-rhamnopyranoside [**15**] (5 mg) identical (<sup>13</sup>C nmr, optical rotation) to an authentic sample.

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#### LITERATURE CITED

1. J. V. Eichholzer, I. A. S. Lewis, J. K. MacLeod, and P. B. Oelrichs, *Tetrahedron*, **37**, 1881 (1981).
2. I. A. S. Lewis, J. K. MacLeod, and P. B. Oelrichs, *Tetrahedron*, **37**, 4305 (1981).
3. M. Klingenberg, M. Appel, and P. B. Oelrichs, *FEBS Lett.*, **189**, 245 (1985).
4. R. Santi and S. Luciani, Eds., "Atractyloside: Chemistry, Biochemistry and Toxicology," Piccin Medical Books, Padua, 1978.
5. K. Heyns and D. Müller, *Tetrahedron*, **21**, 3151 (1965).
6. I. A. S. Lewis and J. K. MacLeod, *Org. Magn. Reson.*, **18**, 138 (1982).
7. C. Laffite, A. M. Nguyen Phuoc Du, F. Winternitz, R. Wylde, and F. Pratiel-Sosa, *Carbohydr. Res.*, **67**, 105 (1978).
8. C. Schöpf and W. Wüst, *Ann.*, **626**, 150 (1959).
9. W. T. Haskins, R. M. Ham, and C. S. Hudson, *J. Am. Chem. Soc.*, **68**, 628 (1946).
10. P. B. Oelrichs, P. J. Vallyly, J. K. MacLeod, and I. A. S. Lewis, *J. Nat. Prod.*, **43**, 414 (1980).

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